# Laser Assisted Root Canal Disinfection-A Review

Tina Puthen Purayil<sup>1,\*</sup>, Arumugam Chakravarthy<sup>2</sup>, Nidambur Vasudev Ballal<sup>3</sup> and Jothi Varghese<sup>4</sup>

<sup>1</sup>Department of Conservative Dentistry and Endodontics, Manipal College of Dental Sciences, Manipal University, Manipal, Karnataka, India

<sup>2</sup>Department of Conservative Dentistry and Endodontics, Sri Ramachandra Dental College, Chennai, India

<sup>3</sup>Department of Conservative Dentistry and Endodontics, Manipal College of Dental Sciences, Manipal University, Manipal, Karnataka, India

<sup>4</sup>Department of Periodontology, Manipal College of Dental Sciences, Manipal University, Manipal, Karnataka, India

**Abstract:** Laser assisted procedures evoked the field of endodontics almost 3-4 decades ago by means of using appropriate energy produced by light for alteration of various hard and soft tissues. During the initial years, the cumbersome laser delivery systems and handling limited its use, despite the purported advantages. However, the last few years, have seen tremendous advancements in terms of tip designs, disinfection agents, photoacoustic streaming etc. This paper has been compiled in an attempt to discuss the effects of photo-activated technology on the root canal system.

Keywords: Laser irradiation, photodynamic therapy, root canal disinfection, smear layer.

#### **1. INTRODUCTION**

The sequel of endodontic treatment is acutely linked to the subsistence of microorganisms in the root canal anatomy. It is well established that microorganisms play a central role in the pathogenesis of pulpoperiapical disease. Endodontic therapy aims to adequately outline the root canal system to achieve disinfection by eliminating bacteria, their toxins, and removing smear layer. In pursuance of achieving a reduction of the microbial count in the root canals, diversified approaches have been suggested involving endodontic preparation techniques, irrigation protocols and use of disinfecting medicaments. Paque et al. [1] have reported that in post mechanical instrumentation, large areas of root canal remain untouched, regardless of whether manual or rotary technique was used. Oguntebi et al. [2] in addition showed that the ability of intracanal medicaments to penetrate into the dentinal tubules is limited and also some of them do not possess a wide spectrum of action against endodontic pathogens. Thus, elimination of microorganisms from the root canals using traditional instrumentation and decontamination techniques remains uncertain. Off late, the use of lasers is increasingly being suggested to attain disinfection of the root canal system and removal of smear layer. The complexities within the

\*Address correspondence to this author at the Department of Conservative Dentistry and Endodontics, Manipal College of Dental sciences, Manipal University, Manipal-576104, Karnataka, India; Tel: -09448332741; Fax: -0820 2922172; E-mail: tina\_p\_p@yahoo.com

root canal harbours locales like secondary canals, anastomoses and deep dentinal tubules which are inaccessible to the conventional disinfecting irrigants and intracanal medicaments. Laser therapy have provided access to these elusive areas and effectively eradicated microorganisms from the deeper reaches of the endodontic reticulation.

### 2. SMEAR LAYER REMOVAL

A residual layered reservoir that contains organic and inorganic debris which is often formed after root canal preparation, with a penetrability of  $40\mu$  in dentin [3]. The elimination of this smear layer has resulted in improved periapical healing.

Various methods have be improvised for its effective elimination and the recent approach rely on laser light when irradiated inside a root canal results in formation of bubbles inside a fluid. This is followed by a phenomenon known as acoustic streaming caused by generation of shock waves due to rapid changes in pressure/amplitude. A forced collapse of bubbles causes implosions that impact on surfaces causing shear forces. In the closed environment of the root canal system these shear forces clear away the smear layer, herniate bacterial cells and disrupt the endodontic surface biofilm thus enhancing their disinfecting and debriding ability [4, 5]. The photothermal and photomechanical actions of laser facilitate smear layer removal. However, since the actions are based on thermal effects, heat generation

and its associated undesirable effects are always a matter of concern. In this regard, it is essential that the appropriate parameters for the selected operation are validated and their safety documented. A quantum of studies has been done to study the various parameters that can be safely employed.

George et al. [6] studied the smear layer removing ability of modified flat and conical tips, with Er:YAG and Er,Cr:YSGG (400µm diameter) laser. Laser activation of irrigants (EDTAC) resulted in sanitised dentinal walls devoid of smear layer. Also, only a 2.5°C increase in temperature was seen when a power of 1 and 0.75W was used. Arslan et al. [7] evaluated the efficacy of 808-nm diode laser agitation of 15% ethylene diaminetetraacetic acid (EDTA) and its effectiveness in smear layer removal. Effective smear layer removal was noted in the apical thirds of root canal following 20 seconds agitation of the irrigant with the diode laser. Hulsman et al. [8] exhibited complete smear layer removal with open dentinal tubules with laser activation of EDTA irrigant without any undesirable phenomena. Intact collagen structure was seen under scanning electron microscope (SEM) in the root dentin suggesting the conservative nature of this treatment modality. De Moor et al. [9] studied the effect of laser activation of irrigants (LAI). They activated 2.5% sodium hypochlorite (NaOCI) with Er,Cr:YSGG laser at 75mJ, 20Hz, 1.5W for 5 seconds with an endodontic (flat end) tip of 200µm diameter. The laser energy energised the irrigants producing photoacoustic streaming. Thermal expansion causing subsequent implosion generated secondary cavitation effect on intracanal fluids. The tip was held stationary 5mm from the apex in the middle third of the canal, without being retracted or moved up and down thus greatly simplifying the procedure. De Moor et al. [4] also compared the efficacy of LAI technique with PUI (Passive ultrasonic irrigation) and CI (Conventional Irrigation) and concluded that laser technique provided superior results with lower irrigation times. Studies by De Groot et al. [5] and Hmud et al. [10] also supported the results by De Moor et al. Macedo et al. [11] stated that laser activation increased potency of NaOCI and increased the consumption of chlorine significantly more compared to PUI or CI.

# 3. DISINFECTION OF ROOT CANAL SYSTEM

Conventionally irrigants have been used to achieve disinfection of root canals with or without mechanical agitation. Laser can be considered as a suitable alternative to achieve endodontic disinfection [12]. Though the bactericidal effects of lasers have been well established but the exact mechanism has not been fully decoded. The antibacterial action depends on the type of laser used. Anti-bacterial effect of Nd:YAG lasers mainly results from its thermal effects, whereas strong water absorption has been suggested for the bactericidal mechanism of action for Er:YAG lasers [13]. Lasers are reported to destroy deeply located bacteria in radicular dentin than traditional chemomechanical methods [14]. A substantial reduction of the bacterial load has been observed when laser is used an adjunct to conventional endodontic treatment [15]. Both near infrared lasers or medium infrared lasers can be used to achieve endodontic disinfection [12, 16].

Lasers of near infrared wavelength have no affinity to hard tissues and thus have no ablative action and are thus is safely employed intraradicularly for disinfection. The root canals are prepared in the conventional way employing mechanical instrumentation until ISO 25/30 apical preparation. Laser irradiation is done at the conclusion of endodontic shaping just prior to obturation as a final means of decontamination. Clinically a fine 200µm diameter endodontic tip is positioned 1mm short of the apical terminus and using a helical movement the tip is retracted coronally at a steady pace in 5 to 10 seconds. The lasing procedure is done in canal filled with an irrigant such as EDTA or NaOCI to avoid undesirable thermal effects. Schoop et al. [13] demonstrated superior efficacy of lasers in disinfection compared to traditional chemical irrigation, in vitro owing to the energy spread of laser irradiation and penetrability into the dentinal wall. The diffusion capacity varied with the wavelength used. A bacterial reduction of 85% at 1mm was demonstrated using Nd:YAG (1064nm) when compared with diode laser(810nm) which exhibited a reduction of 63% at 750µm or less. Diode laser (810nm) when used in combination with endodontic irrigants, EDTA and citric acid brought about a more or less absolute reduction of the bacterial load (99.9%) of E. faecalis in the endodontic system [17].

Erbium lasers possess high affinity to water. They are absorbed primarily on the surface of root canals and exert high bactericidal activity against gram positive (*E. faecalis*) and gram negative *E. coli* bacteria [12]. Lasers possess high bactericidal action owing to their thermal effect and can penetrate deep into the dentinal tubules. Er:YAG laser exerts bactericidal action against *A. naeslundii, E. faecalis, L caesei, P. acnes, F. nucleatum, P. gingivalis* and has been shown to be able to disrupt endodontic biofilm even in the apical third [18]. Moritz *et al.* [19] obtained an almost total eradication (99.64%) of *E. faecalis* using Er:YAG laser following root canal preparation.

Following preparation of the root canal with conventional instrumentation long, thin (200 and 320µm) Erbium laser tip is placed till the working length and retracted using a helical movement over 5 to 10 second interval. The procedure is repeated 3 or 4 times alternating with irrigation with chemical irrigants (NaOCI and/or EDTA) keeping the canal wet while irradiating with laser.

Conventionally mechanical or ultrasonic agitation of endodontic irrigants (NaOCI in combination with chelating agents such as EDTA and citric acid) is carried out to achieve disinfection and cleaning of the endodontic system. Berutti *et al.* [20] have reported action of NaOCI upto 130 $\mu$  depth on the root canal walls. Chen *et al.* [21] have exhibited bactericidal action upto a depth of 300 to 400 $\mu$ m below the lased surface. Bactericidal action results from structural modifications in bacterial cell wall which alters the osmotic gradient, causing cellular distension and finally death.

### 3.1. Dry vs Wet Laser Irradiation

Thermal effects of lasers help kill bacterial cells but also manifests as detrimental changes in the root canal dentinal and adjacent periodontal ligament. Collateral effects of root canal decontamination are seen with both near and medium infrared lasers when used dry. Melting of inorganic dentinal structures seals off tubules. Also recrystallisation bubbles and cracks are evident with partially removed smear layer. To minimize the undesirable thermal effects of laser beam on the dentinal wall laser activation is done in wet canals. The water limits the diffusion of heat onto the dentinal walls and it gets thermally activated either directly by near infrared laser irradiation or indirectly by vaporisation with a medium infrared laser [22, 23, 24].

Yamazaki *et al.* [24] and Kimura *et al.* [25] studied effects of erbium laser and concluded that water is necessary to avoid undesirable morphologic side effects when dry canals are irradiated. Thermal damage and ablation result at the power used for decontamination. Crazing, ledges, zones of superficial melting and smear layer ablation are evident. However, when Erbium laser is irradiated in a wet canal, the thermal damage is minimised and establishes patency of dentinal tubules. Less ablated areas are seen in the more calcified peritubular dentin than inter tubular dentin which is rich in water. Also, following irradiation with erbium laser the smear layer is ablated and mostly eradicated. Moritz *et al.* [19] observed a temperature rise only  $3.5^{\circ}$ C on the periodontal surface when

standardised energy of 100mJ, 15Hz and 1.5W which

### 4. PHOTOACTIVATED DISINFECTION (PAD)

is safely tolerated.

Anti-bacterial effect of the laser used is dependent on its pulse length, fluence, irradiance apart from its wave length [26]. Lasers commonly are used in isolation but can also be used with photosensitizers in a technique called photoactivated disinfection. The mode of action varies completely from that described for lasers used alone. Photodynamic therapy originally was developed as an anticancer therapy. But thereon its application has also been expanded to eradicate bacteria and viruses. The core of this therapy is based on three basic elements: a photosensitizer, a light source and tissue oxygen form. The mechanism of action suggested is due to generation of free radicals. In photoactivated disinfection a photosensitizing compound is topically applied to the hard tissue site to be disinfected and then irradiated with a laser light of a suitable wavelength absorbed by the photosensitizing compound. When a laser of a suitable wavelength (diode) is used, the photosensitizer gets excited, highly cytotoxic singlet oxygen is generated. Localised exposure to light eliminates bacteria whereas the reactive oxygen species act on bacteria and tissue that have selectively taken up the photosensitizer leading to their destruction [27]. A nontoxic photosensitizer is used that is selectively absorbed in a target tissue that responds to a low-intensity light source. In the presence of oxygen, following photo-induced activation, the photosensitizer, produces singlet oxygen and free radicals which damage the bacterial membrane and DNA. The extent of tissue/cell damage depends on the type, dose, incubation time, localization of the photosensitizer, availability of oxygen and also the wavelength of light (nm). A photosensitizer should absorb laser light in the red portion of the visible spectrum or at longer wavelengths in the near infrared region [28].

Bonsor *et al.* [29] have shown PAD and citric acid to be as effective as sodium hypochlorite in disinfection of root canals. PAD also provides a broad-spectrum effect, against some difficult to eradicate organisms [30]. Rovaldi *et al.* [31] have also shown that it is effective even in the presence of blood. Because of its high antibacterial potential PAD has been suggested as an adjunct to conventional endodontic disinfection protocols [32]. Commonly photosensitizer / laser combinations are:

- Tolonium chloride with a 635nm diode laser or 632.8nm helium –neon laser.
- b) Methylene blue with 670nm diode laser.
- c) Aluminum disulfonatedphthalocyanine with a 660nm diode laser.

# 5. PHOTON INDUCED PHOTOACOUSTIC STREAMING (PIPS)

PIPS technique is based upon the interaction of erbium laser and irrigating solutions. It utilises the photoacoustic and photomechanical phenomena exclusively and is not reliant on the thermal action of the laser. PIPS is carried out at subablative intensity with parameters of 20mJ at 15Hz, with 50 microsecond impulses. Each impulse (peak power of 400W) interacts the water molecules with creating successive "shock waves" without the undesirable thermal effects seen with traditional laser techniques. The photoacoustic shockwave generated courses irrigants powerfully throughout the entire root canal system (lateral canals, dentinal tubules and canal anastomoses to the apex) results in high level endodontic debridement and disinfection.

A new tip design with a stripped end is employed in PIPS technique to allow for greater lateral emission of energy compared to the frontal tip. A 600 micron diameter, 9 millimeter long tapered tip with a 3 millimeter stripped end is used. The tip is placed inside the pulp chamber or extended till the coronal 1/3 of root canal and left stationary unlike the LAI technique which the laser needs tip to be placed to within mm from the apical terminus thus greatly simplifying the procedure.

## 5.1. Advantages of Pips

- Effective cleaning and debridement of the root canal system minimising the risk of reinfection.
- Preservation of tooth structure owing to the minimally invasive nature of PIPS.
- Elimination of the risks associated with conventional endodontic preparation techniques like lodging or canal transportation etc.
- Avoidance of unwanted apical extrusion of chemical irrigants since the PIPS tip is only inserted into the pulp chamber or coronal 1/3 of the canal, also there is no risk of tip breakage from curved canals.

Time efficient - Laser activation of chemical agents reduces holding time and saves 20-30 minutes per canal for patient and clinician.

# 6. NEWER TIP DESIGNS

Lasers have a lot of purported advantages, however, there are also several limitations associated with the intracanal use of lasers. Conventionally laser energy is emitted from the end of the optical fiber or the laser guide tip ("end firing tips") and is directed along the root canal and does not necessary spread laterally along the root canal walls [33]. With the conventional end firing tips it is highly improbable to uniformly irradiate the complete root canal surface using a laser [34]. Also there is a potential for thermal damage to the periapical tissues associated with intracanal use of laser [35]. In order to irradiate all areas of the root canal walls and at the same time to ensure the safety of such intracanal usage of lasers various tips were developed.

- Endolase side-firing spiral tip- designed specifically to allow for lateral irradiation of root canal walls. Spiral slit located all along the tip emits the Er:YAG laser irradiation laterally throughout the length of the tip. The tip is sealed at its far end preventing apical irradiation of laser.
- Radial firing tip-passage of laser energy from the tip end towards and through the apical foramen of the tooth causes periodontal damage. A patented 200µm RADIAL FIRING TIPS (Water Lase MD) has been designed for use with an Er:Cr:YSGG laser having a unique beam pattern that does not fire directly into the apex of the root.
- A conical tip which has a fan-shaped emission profile, delivers 80 percent of the energy laterally, and only 20 percent in the forward direction.
- Honeycomb tips with safe ends can activate fluids placed in the root canal and generate shockwaves that are directed onto the walls of the root canals and also into the intricacies of the root canal system. There is very little apical emission, which serves to eliminate the problem of driving fluids beyond the apical foramen. While studying the fluid movements association with laser activation, honeycomb tip design produced agitation driving fluid onto the root

canal walls. In comparison, the conventional plain optical fibers and the conical tips created fluid movement predominantly directed in apical direction [36]. A further advantage of these tip designs is that they are simpler to use in practice because it is no longer necessary for the operator to follow complex sequences of moving and withdrawing the fiber in order to achieve even irradiation of the canal walls.

 Spherical and cylindrical tips with a near-360degree emission profile (isotropic tips) have been used for photoactivated disinfection of root canals using low-intensity visible red light, but the designs are not suited to delivering highintensity pulses in the near (780-1400nm) and mid-infrared (1400-3000nm) range.

# CONCLUSION

With the advancements in laser properties, it has been possible to invade the obscure apical zone of the root canal system thus providing synergistic gains to conventional endodontic therapy. The use of photoinduced disinfection techniques and contemporary tip designs have paved channels to eliminate pathogens and expedite periapical tissue repair, hence creating promising multifaceted applicability.

### REFERENCES

- [1] Paque F, Musch U and Hulsmann M. Comparison of root canal preparation using RaCe and ProTaper rotary Ni-Ti instruments. Int Endod J 2005; 38: 8-16. <u>https://doi.org/10.1111/j.1365-2591.2004.00889.x</u>
- [2] Oguntebi BR. Dentine tubule infection and endodontic therapy implications. Int Endod J 1994; 27: 218-22. <u>https://doi.org/10.1111/j.1365-2591.1994.tb00257.x</u>
- [3] Ruddle C. Cleaning and shaping the root canal system. In:Cohen S, Burns RC, editors. Pathways of the Pulp, ed 8, St Louis, Mosby 2002; p 231-273.
- [4] De Moor RJ, Blanken J, Meire M and Verdaasdonk R. Laser induced explosive vapor and cavitation resulting in effective irrigation of the root canal. Part 2: evaluation of the efficacy. Lasers Surg Med 2009; 41: 520-23. https://doi.org/10.1002/lsm.20797
- [5] de Groot SD, Verhaagen B, Versluis M, Wu MK, Wesselink PR and van der Sluis LW. Laser-activated irrigation within root canals: cleaning efficacy and flow visualization. Int Endod J 2009; 42: 1077-83. <u>https://doi.org/10.1111/j.1365-2591.2009.01634.x</u>
- [6] George R, Meyers IA and Walsh LJ. Laser activation of endodontic irrigants with improved conical laser fiber tips for removing smear layer in the apical third of the root canal. J Endod 2008; 34: 1524-27. https://doi.org/10.1016/j.joen.2008.08.029
- [7] Arslan H, Ayrancı LB, Karatas E, Topçuoğlu HS, Yavuz MS and Kesim B. Effect of agitation of EDTA with 808-nanometer diode laser on removal of smear layer. J Endod 2013; 39: 1589-92. https://doi.org/10.1016/j.joen.2013.07.016

Hülsmann M, Heckendorff M and Lennon A. Chelating agents in root canal treatment: mode of action and indications for their use. Int Endod J 2003; 36: 810-30.

[8]

[9] De Moor RJ, Meire M, Goharkhay K, Moritz A and Vanobbergen J. Efficacy of ultrasonic versus laser-activated irrigation to remove artificially placed dentin debris plugs. J Endod 2010; 36: 1580-83. <u>https://doi.org/10.1016/j.joen.2010.06.007</u>

https://doi.org/10.1111/j.1365-2591.2003.00754.x

- [10] Hmud R, Kahler WA, George R and Walsh LJ. Cavitational effects in aqueous endodontic irrigants generated by nearinfrared lasers. J Endod 2010; 36: 275-78. https://doi.org/10.1016/j.joen.2009.08.012
- [11] Macedo RG, Wesselink PR, Zaccheo F, Fanali D and van der Sluis LWM. Reaction rate of NaOCI in contact with bovine dentine: effect of activation, exposure time, concentration and pH. Int Endod J 2010; 43: 1108-15. <u>https://doi.org/10.1111/j.1365-2591.2010.01785.x</u>
- [12] Gordon W, Atabakhsh VA, Meza F, Doms A, Nissan R, Rizoiu I, et al. The antimicrobial efficacy of the erbium, chromium:yttrium-scandium- gallium-garnet laser with radial emitting tips on root canal dentin walls infected with Enterococcus faecal is. J Am Dent Assoc 2007; 138: 992-02. <u>https://doi.org/10.14219/jada.archive.2007.0297</u>
- [13] Schoop U, Kluger W, Moritz A, Nedjelik N, Georgopoulos A and Sperr W. Bactericidal effect of different laser systems in the deep layers of dentin. Lasers Surg Med 2004; 35: 111-16. https://doi.org/10.1002/lsm.20026
- [14] Klinke T, Klimm W and Gutknecht N. Antibacterial effects of Nd:YAG laser irradiation within root canal dentine. J Clin Laser Med Surg 1997; 15: 29-31.
- [15] Garcez AS, Nu'nez SC, Hamblin MR, Suzuki H and Ribeiro MS. Photodynamic therapy associated with conventional endodontic treatment in patients with antibiotic resistant microflora: a preliminary report. J Endod 2010; 36: 1463-66. <u>https://doi.org/10.1016/j.joen.2010.06.001</u>
- [16] Gutknecht N, Franzen R, Schippers M and Lampert F. Bactericidal effect of a 980-nm diode laser in the root canal wall dentin of bovine teeth. J Clin Laser Med Surg 2004; 22(1): 9-13. https://doi.org/10.1089/104454704773660912
- [17] Schoop U, Moritz A, Kluger W, Patruta S, Goharkhay K, Sperr W, et al. The Er:YAG laser in endodontics: results of an in vitro study. Laser Surg Med 2002; 30: 360-64. https://doi.org/10.1002/lsm.10054
- [18] Noiri Y, Katsumoto T, Azakami H and Ebisu S. Effects of Er:YAG laser irradiation on biofilm-forming bacteria associated with endodontic pathogens in vitro. J Endod 2008; 34: 826-29. https://doi.org/10.1016/j.joen.2008.04.010
- [19] Moritz A, Schoop U, Goharkhay K, Jakolitsch S, Kluger W, Wernisch J, et al. The bactericidal effect of Nd:YAG, Ho:YAG, and Er:YAG laser irradiation in the root canal: An in vitro comparison. J Clin Laser Med Surg 1999; 17: 161-64.
- [20] Berutti E, Marini R and Angeretti A. Penetration ability of different irrigants into dentinal tubules. J Endod1997; 23: 725-27. https://doi.org/10.1016/S0099-2399(97)80342-1
- [21] Chen WH. Laser in root canal therapy. J Indiana Dent Assoc2003; 81: 20-3.
- [22] da Costa Ribeiro A, Nogueira GEC, Antoniazzi JH, Moritz A and Zezell DM. Effects of diode laser (810nm) irradiation on root canal walls: Thermographic and morphological studies. J Endod 2007; 33: 252-55. https://doi.org/10.1016/j.joen.2006.09.002
- [23] He H, Yu J, Song Y, Lu S, Liu H and Liu L. Thermal and morphological effects of the pulsed Nd:YAG laser on root canal surfaces. Photomed Laser Surg 2009; 27: 235-40. <u>https://doi.org/10.1089/pho.2008.2244</u>

- [24] Yamazaki R, Goya C, Yu D-G, Kimura Y and Matsumoto K. Effects of erbium, chromium:YSGG laser irradiation on root canal walls: A scanning electron microscopic and thermographic study. J Endod 2001; 27: 9-12. https://doi.org/10.1097/00004770-200101000-00003
- [25] Kimura Y, Yu DG, Kinoshita J, Hossain M, Yokoyama K, Murakam Y, et al. Effects of erbium, chromium: YSGG laser irradiation on root surface: Morphological and atomic analytical studies. J Clin Laser Med Surg 2001; 19: 69-72. <u>https://doi.org/10.1089/104454701750285386</u>
- [26] Meire MA, De Prijck K, Coenye T, Nelis HJ and De Moor RJG. Effectiveness of different laser systems to kill Enterococcus faecalis in aqueous suspension and in an infected tooth model. Int Endod J 2009; 42: 351-59. <u>https://doi.org/10.1111/j.1365-2591.2008.01532.x</u>
- [27] Dougherty TJ, Gomer CJ, Henderson BW, Jori G, Kessel D, Korbelik M, et al. Photodynamic therapy. J Natl Cancer Inst 1998; 90: 889-905. https://doi.org/10.1093/inci/90.12.889
- [28] Komerik N and Mac Robert AJ. Photodynamic therapy as an alternative antimicrobial modality for oral infections. J Environ Pathol Toxicol Oncol 2006; 25: 487-504. https://doi.org/10.1615/JEnvironPatholToxicolOncol.v25.i1-2.310
- [29] Bonsor SJ, Nichol R, Reid TMS and Pearson GJ. Microbiological evaluation of photo-activated disinfection in endodontics. Br Dent J 2006; 200: 337-41. <u>https://doi.org/10.1038/sj.bdj.4813371</u>

Received on 30-10-2016

Accepted on 01-12-2016

Published on 20-12-2016

DOI: http://dx.doi.org/10.12974/2311-8695.2016.04.02.1

© 2016 Purayil et al.; Licensee Savvy Science Publisher.

This is an open access article licensed under the terms of the Creative Commons Attribution Non-Commercial License (<u>http://creativecommons.org/licenses/by-nc/3.0/</u>) which permits unrestricted, non-commercial use, distribution and reproduction in any medium, provided the work is properly cited.

- Purayil et al.
- [30] Bergmans L, Moisiadis P, Huybrechts B, Van Meerbeek B, Quirynen M and Lamberchts P. Effect of photo activated disinfection on endodontic pathogens ex vivo. Int Endod J 2008; 41: 227-39. <u>https://doi.org/10.1111/j.1365-2591.2007.01344.x</u>

[31] Rovaldi CR, Pievsky A, Sole NA, Friden PM, Rothstein DM and Spacciapoli P. Photoactive Porphyrin Derivative with Broad-Spectrum Activity against Oral Pathogens In vitro. Antimicrob Agents Chemother 2000; 44: 3364-7. <u>https://doi.org/10.1128/AAC.44.12.3364-3367.2000</u>

- [32] Mohan D, Maruthingal S, Indira R, Divakar DD, Al Kheraif AA, Ramakrishnaiah R, et al. Photo activated disinfection (PAD) of dental root canal system - An ex-vivo study. Saudi J Biol Sci 2016; 23: 122-7. <u>https://doi.org/10.1016/j.sjbs.2015.01.013</u>
- [33] Stabholz A, Zeltzser R, Sela M, Peretz B, Moshonov J, Ziskind D, et al. The use of lasers in dentistry: principles of operation and clinical applications. Compend Contin Educ Dent 2003; 24:935-48.
- [34] Goodis HE. Commentary on: Filling root canals in three dimensions. J Endod 2006; 32: 279-80. https://doi.org/10.1016/j.joen.2006.02.029
- [35] Kimura Y, Wilder-Smith P and Matsumoto K. Lasers in endodontics: a review. Int Endod J 2000; 33: 173-85. https://doi.org/10.1046/j.1365-2591.2000.00280.x
- [36] George R and Walsh LJ. Performance assessment of novel side firing safe tips for endodontic applications. J Biomed Opt 2011; 16: 048004. <u>https://doi.org/10.1117/1.3563637</u>